

Diterpenes from *Sideritis infernalis* and *S. candicans*

Braulio M. Fraga^a, Carlo Bressa^a, Concepción Fernández^a, Pedro González^b, Ricardo Guillermo^b, and Melchor G. Hernández^a

^a Instituto de Productos Naturales y Agrobiología, CSIC, Avda. Astrofísico F. Sánchez 3, 38206-La Laguna, Tenerife, Canary Islands, Spain

^b Instituto de Bioorgánica "Antonio González", Universidad de La Laguna, Tenerife, Spain

Reprint requests to Prof. Dr. B. M. Fraga. Fax: 34-922260135. E-mail: bmfraga@ipna.csic.es

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A phytochemical study of *Sideritis infernalis* led to the isolation of the new *nor*-diterpene adejone (17-*nor*-7 α ,18-dihydroxy-*ent*-kaur-16-one). The biosynthesis of this compound implies the decarboxylation of an epoxy-acid as the last step. In addition, three diterpenes with an *ent*-kaurene skeleton, episideridiol, candicandiol 7 α -monoacetate and candidiol 15 α -monoacetate, have been isolated from *S. candicans* for the first time in nature.

Key words: *Sideritis*, Diterpenoids, Adejone, Episideridiol

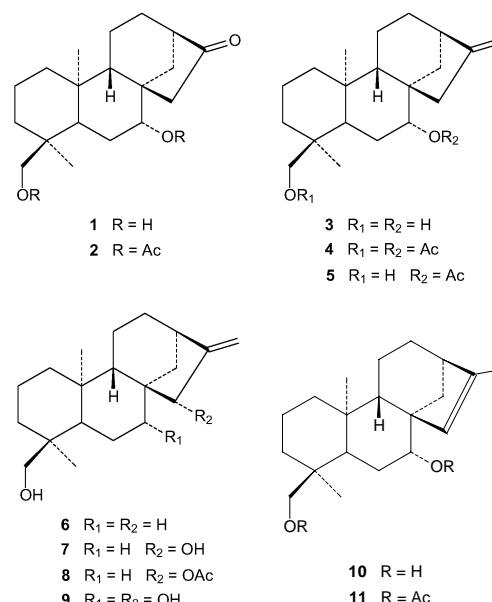
Introduction

The genus *Sideritis* (Lamiaceae) is represented in the Canary Islands by 22 species [1]. We have been interested in their phytochemistry and phylogeny for several years [2, 3].

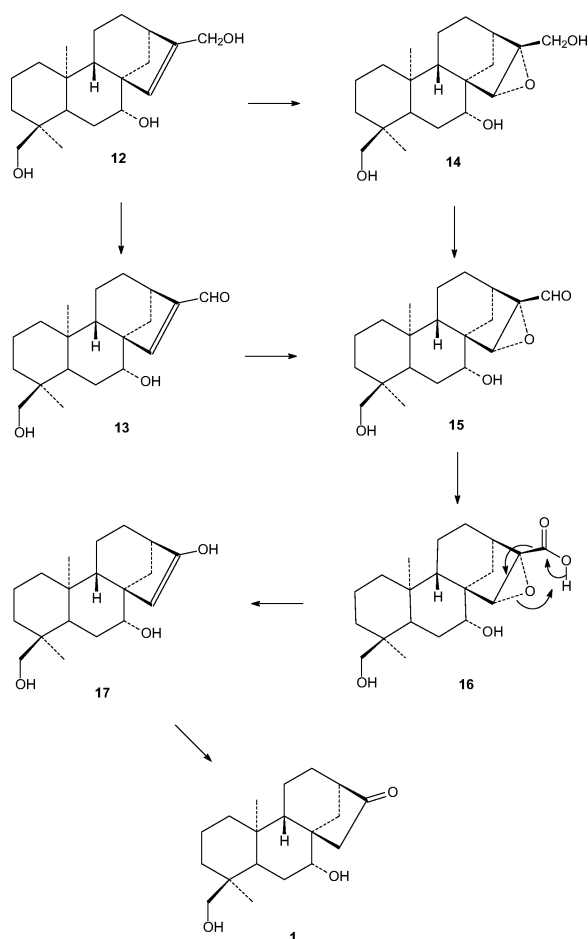
In a continuation of this work we have now completed earlier studies of *S. infernalis* Bolle [4, 5] and reinvestigated *S. candicans* Ait [*S. oroteneriffae* L. Negrín et P. Pérez] [6]. Both species are endemic to the island of Tenerife. We have followed Sventenius's work [7] naming the species of this genus. However, we include the actual botanical name between square brackets. The whole plant of *S. candicans* has been used in Tenerife as an anticatarrhal and stimulant of the circulatory system [8].

Results and Discussion

The diterpenes candicandiol (3), candol B (6), candidiol (7), canditriol (9), sinfernal (12), sinfernal (13), epoxysinfernal (14), the 7-epimer of sinfernal and the flavanones 5-hydroxy-7,4'-dimethoxyflavanone and 5-hydroxy-7,3',4'-trimethoxyflavanone had been obtained from an extract of *S. infernalis* Bolle [4, 5]. Now, in this work, we give an account of other compounds later identified in this extract. One of them was a new *nor*-diterpene, which we have named adejone (17-*nor*-7 α ,18-dihydroxy-*ent*-kaur-16-one) (1). This compound was obtained in the form of its diacetate 2 by acetylation and chromatog-



raphy of fractions containing it. Its high-resolution MS was in accordance with the structural formula C₂₃H₃₃O₅. The ¹H NMR spectrum showed resonances of two angular methyl groups, the H-18 as a pair of doublets at δ = 3.60 and 3.85 (J = 11.0 Hz) and the axial geminal hydrogen of the C-7 acetate at δ = 4.74 (dd, J = 11.6 and 4.2 Hz). The ¹³C NMR spectrum showed the resonances of C-7 and C-18 at δ = 75.8 and 72.0, respectively, while the C-6-oxo group appears at δ = 220.9. Both spectra were unambigu-



Scheme 1.

ously assigned using 2D NMR data (COSY, HSQC, HMBC and NOESY). In the HMBC spectrum correlations were observed of H-5 with C-1, C-7, C-19 and C-20; H-7 with C-5, C-6, C-14 and C-15; H-9 with C-1, C-12 and C-14; H-15 with C-7, C-9, C-13 and C-16; H-18 with C-3, C-4, C-5 and C-19; H-19 with C-3, C-4, C-5 and C-18; and H-20 with C-1, C-5, C-9 and C-10.

Hydrolysis of adejone diacetate (**2**) with methanolic potassium hydroxide (3%) at r.t. led to adejone (**1**), which is the natural diterpene formed in the plant. Their ^1H , ^{13}C and 2D NMR spectra were also in accordance with the assigned structures. Chemical confirmation of the structure of adejone was obtained by ozonolysis of candicandiol diacetate (**4**), which gave a product identical with adejone diacetate (**2**). This is the first time that a 16-oxo-17-*nor*-kaurane has been isolated as a natural product. The diterpene 17-*nor*-

7 α ,19-dihydroxy-*ent*-kaur-16-one, the 4-epimer of **1**, had been prepared by 7 α -hydroxylation of synthetic 17-*nor*-19-hydroxy-*ent*-kaur-16-one with the fungus *Rhizopus nigricans* [9].

A biogenetic route from sinferol (**12**) account for the biosynthesis of adejone (**1**) (Scheme 1). The diterpenes sinferol (**12**), sinferval (**13**) and epoxysinferol (**14**) involved in this route have also been isolated from this plant [4]. The same enzyme probably produces oxidations that transform the C-17 alcohol of sinferol (**12**) into the corresponding acid **16**. The main step is a glycidic acid type decarboxylation of **16** to give the enol **17** [10]. In this reaction, decarboxylation and epoxide ring opening occur simultaneously to form the enol, which tautomerizes giving adejone (**1**). The decarboxylation of an epoxy-acid is a very rare step in the formation of a natural product.

Other compounds now identified in this species were the methyl ester of *p*-methoxycinnamic acid and a mixture of the sterols β -sitosterol, stigmasterol and campesterol.

We have now reinvestigated *S. candicans* Ait. In a previous work we had isolated from this species the diterpenes *ent*-kaur-16-ene, dehydroabietane, epicandicandiol 7 β -monoacetate, trachinodiol 7 β -monoacetate, candicandiol (**3**) and candidiol (**7**), the sterols β -sitosterol, stigmasterol and campesterol, and the triterpenes squalene, glutinol, and ursolic and oleanolic acids [6]. Later, we indicated that the triterpene glutinol had been misidentified as a component of several species of *Sideritis*, and recognized it as its isomer rhoiptelenol [3].

We have now assigned the structure **10** to the diterpene episideridiol, which was obtained as its diacetate **11** by acetylation and chromatography of fractions containing it. Its ^1H NMR spectrum showed signals of three methyls, one at $\delta = 1.67$ (d, $J = 1.5$ Hz) placed over a double bond, two acetates, a vinylic proton at $\delta = 5.10$ (br s) and the two hydrogens of an acetylated primary alcohol at $\delta = 3.60$ and 3.80 (d, $J = 11.1$ Hz), which was assigned to C-18 considering these chemical shifts [11]. The structure was confirmed, and unambiguously assigned by the ^1H and ^{13}C NMR spectra, considering 2D NMR data (HMQC, HMBC, COSY and NOESY). Basic hydrolysis of **11** led to **10**, which is the natural product found in the plant. This is the first time that episideridiol (**10**) is found in nature. It had been synthesized from its 7-epimer sideridiol [12].

Another two compounds that were isolated for the first time from a natural product were candican-diol 7 α -monoacetate (**5**) (30 mg) and candidiol 15 α -monoacetate (**8**) (110 mg). These diterpenes had been prepared from candicandiol (**3**) [13] and candidiol (**7**) [14], respectively, and a direct comparison of these monoacetates, and the corresponding diacetates, confirmed their structures.

Other compounds now identified for the first time in this plant were 2 β -hydroxy-*ent*-13-*epi*-manoyl oxide [15], 16 α ,18-dihydroxy-*ent*-atisane [16], sinfer-nol (**12**), sinfernal (**13**) [4], sidendrodiol [17] and vierol [18]. Adejone (**1**) and epoxysinfernol (**15**) (see above) were tentatively identified in impure chromatographic fractions containing sinfernal (**13**) as the main compound.

We have shown that the Macaronesian *Sideritis* may be classified phytochemically into three groups: *Group 1*, species that contain pentacyclic triterpenes, but not diterpenes, *group 2*, characterized by possessing bicyclic diterpenes and abundance of flavones, and *group 3*, species containing tetra- and/or penta-cyclic diterpenes [2, 6]. Recent morphological studies have shown that these phytochemical groups coincide with the sections *Cretica*, *Empedocleopsis* and *Marrubistrum*, respectively [1].

In accordance with their content in diterpenes, *S. infernalis* Bolle and *S. candicans* Ait [*S. orotenerif-fae* L. Negrín et P. Pérez] belong to the third group. Moreover, these two species, together with *S. bol-leana* Bornm. [*S. barbellata* Mend.-Heuer emend., P. Pérez et L. Negrin] [6], *S. dendrochahorra* Bolle [6, 17] and *S. ferrensis* P. Pérez et L. Negrin [19], are characterized by containing the diterpene candicandiol (**3**). In consequence, from the phytochemical view-point, they can form a new subgroup within our third group of *Sideritis*. Morphologically, three of these species, *S. bolleana*, *S. dendrochahorra* and *S. ferrensis*, belong to the subsection *Massoniana* (Christ) Svent [1].

Experimental Section

General experimental procedures

^1H and ^{13}C NMR spectra were recorded in CDCl_3 at 500.13 and 125.77 MHz, respectively, with a Bruker AMX2-500 spectrometer. Chemical shifts are given in ppm (δ). Mass spectra were taken at 70 eV (probe) in a Micro-mass Autospec spectrometer. Dry column chromatographies were made on silica gel (Merck 0.02–0.063 mm).

C atom	1	2	10	11
1	39.7	39.6	39.7	39.7
2	17.8	17.6	17.8	17.4
3	35.0	35.6	35.0	35.7
4	37.4	36.2	37.4	36.3
5	45.8	46.3	45.7	46.6
6	28.4	24.5	27.5	25.2
7	74.1	75.8	75.0	76.0
8	47.9	46.3	55.3	53.7
9	54.0	54.0	47.9	48.1
10	39.2	39.1	38.9	38.9
11	18.3	18.2	18.4	18.1
12	30.0	29.5	25.2	24.9
13	47.2	46.8	44.0	43.4
14	28.3	29.5	34.6	35.6
15	49.7	49.1	131.9	132.0
16	222.2	220.9	145.7	143.2
17	–	–	15.5	15.3
18	71.6	72.0	71.7	78.3
19	17.6	17.5	18.3	17.7
20	18.6	18.5	17.4	18.4

Table 1. ^{13}C NMR data (δ , in ppm) of compounds **1**, **2**, **10** and **11**.

Isolation of compounds from *S. infernalis*

We have described the isolation of several diterpenes [4] and flavanones [5] from an extract of *S. infernalis* Bolle. Continuing with the study of the components of this extract, we isolated a mixture of β -sitosterol, stigmasterol and campesterol (130 mg), adejone (**1**) and *p*-methoxycinnamic acid methyl ester (130 mg). Adejone (**1**) was obtained in form of the diacetate **2** (11 mg) by acetylation of several fractions (240 mg) of the main chromatography, which contained also candicandiol (**3**) and candidiol (**7**).

Adejone (**1**)

^1H NMR (CDCl_3 , 500 MHz): δ = 0.77 (1 H, td, J = 13.0 and 3.9 Hz, H-1 β), 0.78 (3 H, s, H-19), 1.13 (3 H, s, H-20), 1.21 (1 H, d, J = 8 Hz, H-9), 1.28 (1 H, dt, J = 13.5 and 3.0 Hz, H-3 α), 1.30 (1 H, dd, J = 12.5 and 1.5 Hz, H-5), 1.43 (1 H, td, J = 13.5 and 4.4 Hz, H-3 β), 1.48 (1 H, t, J = 12.5 Hz, H-6 α), 1.73 (1 H, ddd, J = 12.5, 4.0 and 1.5 Hz, H-6 α), 1.89 (1 H, dd, J = 12.0 and 3.5 Hz, H-14), 2.00 (1 H, dd, J = 12.0 and 5.0 Hz, H-14), 2.44 (1 H, br d, J = 3.5 Hz, H-13), 2.64 (1 H, dd, J = 18 and 1.2 Hz, H-15), 3.11 and 3.44 (each 1 H, d, J = 11 Hz, H-18), 3.59 (1 H, dd, J = 12.0 and 4 Hz, H-7). – ^{13}C NMR: see Table 1. – EIMS: m/z (%) = 306 (8) [M] $^+$, 275 (100), 257 (47), 193 (8), 179 (16), 175 (3), 161 (4), 147 (5), 145 (4), 133 (7), 123 (32). – HREIMS: m/z = 306.2194 (calcd. 306.2195 for $\text{C}_{19}\text{H}_{30}\text{O}_3$, [M] $^+$).

Diacetate **2**

^1H NMR (CDCl_3 , 500 MHz): δ = 0.81 (1 H, td, J = 12.8 and 3.9 Hz, H-1 β), 0.84 (3 H, s, H-19), 1.16 (3 H, s, H-20), 1.27 (1 H, d, J = 8 Hz, H-9), 1.35 (1 H, dd, J = 11.5 and 1.4 Hz, H-5), 1.42 (1 H, td, J = 13.3 and 4.4 Hz, H-3 β),

1.80 (1 H, m, H-1 α), 1.87 (1 H, dd, J = 18.6 and 3.1 Hz, H-15), 1.98 (1 H, dd, J = 12.0 and 5.0 Hz, H-14), 2.02 (1 H, dd, J = 12.0 and 4.0 Hz, H-14), 2.05 and 2.10 (each 3 H, s, OAc), 2.20 (1 H, dd, J = 18.6 and 1.1 Hz, H-15), 2.44 (1 H, br s, H-13), 3.60 and 3.85 (each 1 H, d, J = 11.0 Hz, H-18), 4.74 (1 H, dd, J = 11.6 and 4.2 Hz, H-7). – ^{13}C NMR: see Table 1. – EIMS: m/z (%) = 390 (3) $[\text{M}]^+$, 330 (25), 275 (40), 270 (97), 257 (100), 255 (58), 241 (16), 227 (9), 214 (14), 201 (12), 187 (11), 175 (15). – HREIMS: m/z = 390.2407 (calcd. 390.2406 for $\text{C}_{23}\text{H}_{33}\text{O}_5$, $[\text{M}]^+$).

Ozonolysis of candicandiol diacetate (4)

The diacetate **4** (12 mg) was dissolved in dichloromethane (3 mL) and cooled to -78°C . A stream of ozone was bubbled through the solution for 20 min, until it turned to a light blue color. Excess of ozone was removed by bubbling nitrogen through the solution. Triphenylphosphine (16 mg) was added, and the solution was stirred for 1 h at r. t. Usual work up, extraction with EtOAc, and purification by column chromatography afforded adejone diacetate (**2**).

Isolation of compounds from *S. candicans*

The aerial parts of *S. candicans* Ait (*S. oroteneriffae* L. Negrín et P. Pérez) (4.1 kg) were collected in May at the higher parts of Arafo, near of the end of the road from Arafo to La Cumbre (TF-523) on Tenerife Island. The plant was identified by Prof. Pedro Pérez de Paz, Botanical Department, University of La Laguna (Tenerife). A general description of the procedure to isolate the compounds of *Sideritis* species has been published previously [6]. The following substances were isolated: Candol B (**6**) (30 mg), rhoiptelanol (59 mg), a mixture of β -sitosterol, stigmasterol and campesterol (1.3 g), 2 β -hydroxy-*ent*-13-*epi*-manoyl-oxide, trachinodiol 7 β -monoacetate, candidiol 15 α -monoacetate (**8**), candicandiol 7 α -monoacetate (**5**) and epicandicandiol 7 β -monoacetate (obtained by acetylation as their corresponding diacetates, the yields being 21, 480, 30, 110 and

360 mg, respectively), oleanolic and ursolic acids (8 mg), sinfernal (**13**), sinfernol (**12**), 16 α ,18-dihydroxy-*ent*-atisane, sidendrodiol, vierol and episideridiol (**10**) (identified as their corresponding diacetates, the yields being 28, 2, 8, 6, 5 and 20 mg, respectively), candidiol (**7**) (505 mg) and candicandiol (**3**) (620 mg). Adejone (**1**) and epoxysinfernal (**15**) were also identified, as traces, in impure fractions containing sinfernal (**13**) as the main compound.

Episideridiol (10)

^1H NMR (CDCl_3 , 500 MHz): δ = 0.74 (3 H, s, H-19), 0.99 (1 H, d, J = 7.8 Hz, H-9), 1.06 (3 H, s, H-20), 1.17 (1 H, dd, J = 12.2 and 1.7 Hz, H-5), 1.71 (3 H, s, H-17), 1.78 (1 H, dt, J = 12.9 and 3.1 Hz, H-1 α), 2.38 (1 H, br s, H-13), 3.08 and 3.42 (each 1 H, d, J = 10.8 Hz, H-18), 3.65 (1 H, dd, J = 11.3 and 4.2 Hz, H-7), 5.12 (1 H, br s, H-15). – ^{13}C NMR: see Table 1. – EIMS: m/z (%) = 304 (100) $[\text{M}]^+$, 286 (10), 268 (7), 255 (18), 164 (10), 147 (9), 131 (8), 123 (38). – HRMS: m/z = 304.2439 (calcd. 304.2402 for $\text{C}_{20}\text{H}_{32}\text{O}_2$, $[\text{M}]^+$).

Diacetate (11)

^1H NMR (CDCl_3 , 500 MHz): δ = 0.73 (1 H, td, J = 12.9 and 4.2 Hz, H-1 β), 0.80 (3 H, s, H-19), 0.99 (1 H, d, J = 7.6 Hz, H-9), 1.08 (3 H, s, H-20), 1.21 (1 H, dd, J = 12.1 and 1.6 Hz, H-5), 1.67 (3 H, d, J = 1.5 Hz, H-17), 2.00 and 2.08 (each 3 H, s, OAc), 2.31 (1 H, br s, H-13), 3.60 and 3.80 (each 1 H, d, J = 11.1 Hz, H-18), 4.86 (1 H, dd, J = 11.3 and 4.2 Hz, H-7), 5.10 (1 H, br s, H-15). – ^{13}C NMR: see Table 1. – EIMS m/z (%) = 388 (11) $[\text{M}]^+$, 346 (21), 328 (13), 313 (7), 268 (27), 255 (31), 240 (15), 225 (14), 197 (6), 185 (11), 171 (7). – HREIMS m/z = 388.2596 (calcd. for 388.2614 $\text{C}_{24}\text{H}_{36}\text{O}_4$, $[\text{M}]^+$).

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- [1] P. Pérez de Paz, L. Negrín-Sosa, *Revisión taxonómica de Sideritis L. Subgénero Marrubiastrum (Moench) Mend.-Heur., Phanerogamarum monographiae*, J. Cramer, Berlin **1992**.
- [2] B. M. Fraga, *Instituto de Estudios Canarios. 50 Aniversario*, Cabildo Insular, Tenerife **1982**, Vol. 1, pp. 115–135.
- [3] B. M. Fraga, M. Reina, J. G. Luis, M. L. Rodríguez, *Z. Naturforsch.* **2003**, 58c, 621–625.
- [4] C. Fernández, B. M. Fraga, M. G. Hernández, *Phytochemistry* **1986**, 25, 2573–2576.
- [5] C. Fernández, B. M. Fraga, M. G. Hernández, *J. Nat. Prod.* **1988**, 51, 591–593.
- [6] A. G. González, B. M. Fraga, M. G. Hernández, J. G. Luis, F. Larruga, *Biochem. System. Ecol.* **1979**, 7, 115–120.
- [7] E. R. Sventenius, *Collectanea Botanica* **1968**, 7, 1121–1157.
- [8] P. Pérez de Paz, C. Hernández-Padrón, *Plantas medicinales o útiles en la flora canaria*, F. Lemus, La Laguna, Spain, **1999**.
- [9] E. L. Ghisalberti, P. R. Jefferies, M. A. Sefton, P. N. Sheppard, *Tetrahedron* **1977**, 33, 2451–2436.
- [10] S. P. Singh, J. Kagan, *J. Org. Chem.* **1970**, 35, 2203–2207.

- [11] P. R. Jefferies, R. W. Retallac, *Austr. J. Chem.* **1968**, *21*, 2085–2093.
- [12] F. Piozzi, P. Venturella, A. Bellino, M. L. Marino, P. Salvadori, *J. Chem. Soc., Perkin Trans. I* **1972**, 759–762.
- [13] B. M. Fraga, P. González, M. G. Hernández, S. Suárez, *Tetrahedron* **2005**, *61*, 5623–5632.
- [14] B. M. Fraga, M. G. Hernández, P. González, *Phytochemistry* **1992**, *31*, 3845–3849.
- [15] E. Sezik, N. Ezer, J. A. Hueso-Rodríguez, B. Rodríguez, *Phytochemistry* **1985**, *24*, 2739–2740.
- [16] M. P. L. Moraes, N. F. Roque, *Phytochemistry* **1988**, *27*, 3205–3208.
- [17] B. M. Fraga, M. G. Hernández, C. Fernández, J. M. Arteaga, *Phytochemistry* **1987**, *26*, 775–777.
- [18] A. G. González, B. M. Fraga, M. G. Hernández, J. G. Luis, *Tetrahedron* **1973**, *29*, 561–563.
- [19] B. M. Fraga, M. G. Hernández, J. M. H. Santana, J. M. Arteaga, *Phytochemistry* **1991**, *30*, 913–915.